RESEARCH

HISTOPATHOLOGICAL EFFECTS OF LOCAL AND SYSTEMIC OZONE TREATMENT IN EXPERIMENTAL TYMPANIC MEMBRANE PERFORATIONS

Metin YILDIRIM¹, MD; D Şahin ÖĞREDEN¹, MD; D Ümit TAŞKIN¹, MD Alper TABARU¹, MD; D Funda KAYA EMRE², MD; D Mehmet Faruk OKTAY¹ MD

¹Bağcılar Eğitim Araştırma Hastanesi, KBB, İstanbul, Turkey ²Bağcılar Eğitim Araştırma Hastanesi, Patoloji, İstanbul, Turkey

SUMMARY

Aims: To investigate histopathological effects of local and systemic ozone application in experimental tympanic membrane perforations.

Methods: Traumatic perforation was created by using a 21 gauge syringe needle under the microscope and then animals were then divided into 4 equal groups. The first group was the control group and it healed spontaneously. 0.5 mL local ozone oil was dropped to external ear in second group; 1.0 mL (1 μ g / mL) ozone gas was given by rectal way in third group; 0.5 mL local ozone oil and 1.0 mL (1 μ g / mL) rectal ozone gas was given in fourth group for 10 days. On day 11th, the tympanic membranes of the rats were excised and examined for inflammation, fibroblast proliferation, vascular density, collagenization and epithelization.

Results: It was observed that fibroblast proliferation, collagenization and epithelization scores of local, local+rectal, rectal ozone treatment groups were more significant compared to the control group (p<0.05).

Conclusion: This study showed the contribitions of local and systemic ozone treatment to wound healing by increasing inflammation, fibroblast proliferation, collagenization, vascular density and epithelization in experimental tympanic membrane perforations of rats.

Keywords: Trauma, tympanic membrane perforation, wounds, rats, ozone

DENEYSEL TİMPANİK MEMBRAN PERFORASYONLARINDA LOKAL VE SİSTEMİK OZON TEDAVİSINİN HİSTOPATOLOJİK ETKİLERİ

ÖZET

Amaç: Lokal ve sistemik ozon uygulamasının deneysel timpanik membran perforasyonları üzerindeki histopatolojik etkilerini araştırmak.

Yöntemler: Mikroskop altında 21 gauge şırınga iğnesi kullanılarak travmatik perforasyon oluşturuldu ve sonra hayvanlar 4 eşit gruba ayrıldı. İlk grup kontrol grubuydu ve kendiliğinden iyileşmeye bırakıldı. İkinci grupta dış kulak yoluna lokal 0.5 mL ozon yağı ; üçüncü grupta rektal yolla 1.0 mL (1 μ g / mL) ozon gazı; dördüncü grupta 0,5 mL lokal ozon yağı ve 1,0 mL rektal ozon gazı (1 μ g / mL) 10 gün boyunca verildi. 11. günde, sıçanların timpanik membranları çıkarıldı ve inflamasyon, fibroblast proliferasyonu, vasküler yoğunluk, kollajenizasyon ve epitelizasyon açısından incelendi.

Bulgular: Fibroblast proliferasyonu, kollajenizasyon ve epitelizasyon skorlarının lokal, lokal + rektal ve rektal ozon tedavi gruplarında kontrol grubuna kıyasla istatiksel açıdan anlamlı olacak şekilde daha yüksek olduğu gözlendi (p < 0.05).

Sonuç: Bu çalışma, deneysel timpanik membran perforasyonlarında lokal ve sistemik ozon tedavisinin, inflamasyon, fibroblast proliferasyonu, kollajenizasyon, damar yoğunluğu ve epitelizasyonu artırarak yara iyileşmesine katkılarını göstermiştir.

Anahtar Sözcükler: Travma, timpanik membran perforasyonu, yara, sıçan, ozon

INTRODUCTION

Tympanic membrane (TM) perforation is clinically common condition of people but the real incidence is unknown. The estimated incidence is reported below 1% all around the

Corresponding Author: Metin YILDIRIM MD Bağcılar Eğitim Araştırma Hastanesi, KBB, İstanbul, Turkey, E-mail: dr.metinyldrm@gmail.com

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Small traumatic tympanic membrane perforations usually heal spontaneously. In some of these cases, the perforation doesn't close or partially closes with a thin membrane. However, many factors can lead to chronic perforation. In this situations, regeneration of the membrane is incompleted. Histologically, the squamous epithelium migrates through the perforation margin to the inner layer of the membrane. Theoretically, epithelialization of the wound edge stops spontaneous healing. Granulocyte Colony Stimulating Factor (G-CSF), Hepatocyte Growth Factor (HGF), Epidermal Growth Factor (EGF), Gelfoam, Saline irrigation, Ascorbic acid (C vit), Hyaluronic acid were used to accelerate TM repair in experimental studies²⁻⁵.

Previous studies showed ozone treatment accelerates wound healing by increasing growth factors such as VEGF, TGF-B and PDGF. In this study, we thought ozone therapy would increase the wound healing at a cellular level in traumatic tympanic membrane perforations.

MATERIAL and METHODS

This study started after confirmation of Experimental Animal Ethics Board dated on 26.01.2016 and numbered 2016/04. 20 healthy Wistar Hannover genus and male rats about 335 Gr (250-450) weight were used in the study. The rats lived at 16-21C temperature, 50% moisture and fed with Pellet. Our study complied on the part of animal care of Helsinki declaration.

Ketamine hydrochloride (Ketalar; Pfizer Turkey) 60 Drug, Istanbul, mg / kg intramusculer (i.m) and xylazine hydrochloride (Rompun; Bayer Healthcare, Kansas, USA) 12 mg / kg / intramusculer (i.m) were used for anesthesia of rats prior to myringotomy. Then, we examined the 40 ears of 20 rats on microscope. We removed debris and cerumen from the external auditory canal. Healty tympanic membrane images were detected in all rats. Traumatic perforation approximately 1 mm wide was created by using a 21 gauge syringe needle through anterior quadrant of both tympanic membranes of the rats. Then, animals were divided into 4 equal groups. The first group the control group and it healed was spontaneously. 0.5 mL local ozone oil (Arozon ozonated olive oil; Ar-Tekin Drug, Çankırı, Turkey) was dropped to external auditory canal in second group; 1.0 mL (1 μ g / mL) ozone gas was given by rectal way in third group; 0.5 mL local ozone and 1.0 mL (1 μ g / mL) rectal ozone gas were given in fourth group for 10 days. On first day of treatment, one of the rats in local ozone group; on second day of treatment, one rat in rectal group and one rat in rectal+local ozone groups were died and seventeen rats were treated completely. On the 11th day of the study, the animals were sacrified after injection of 80 mg / kg / intramusculer (i.m) ketamine hydrochloride and 20 mg / k.g / i.m. xylazine hydrochloride. Subsequently, temporal bone dissection of the rats was carried out for histopathological examination.

2 of the 10 temporal bones in the 5 control group rats, 2 of the 8 temporal bones in 4 local ozone group rats, 1 of the 8 temporal bones in 4 rectal ozone group rats and 1 of the 8 temporal bones in the 4 rectal + ozone group rats were excluded from the study since tympanic membranes were damaged during dissection, 28 tympanic membranes with tympanic bones could be excised. Then, the tissues were placed in a 10% formaldehyde solution and sent to the pathology. Histopathological examination of tissue specimens were performed without knowledge the test groups. Vascularization fibroblast (vessel density), proliferation, collagenization and epithelialization scores were evaluated as 0: none 1: mild 2: middle 3: severe. Inflammation severity scoring system of Hooker et al was used as 0: no inflammation, 1: rare lymphocytes and plasma cells, 2: increased lymphocyte, neutrophil and plasma cells, 3: a large number of mixed inflammatory cells and micro abscesses ⁶.

Statistical analyzes were performed using the NCSS (Number Cruncher Statistical System) 2007 Statistical Software (Utah, USA) package program. Descriptive statistical methods (mean, standard deviation, median and interquartile range), Kruskal Wallis test in group comparisons, Dunn's multiple comparison test in subgroup comparisons were used to evaluate the data. The results were evaluated at a significance level of p < 0.05.

RESULTS

İnflamation severity

The mean inflammation scores of control, local, rectal and local+rectal treatment groups were orderly 1.38 ± 0.52 , 2 ± 0 , 1 ± 0 , 2.14 ± 0.38 (Table 1). Inflammation values of local + rectal ozone therapy group were found to be significantly higher than the control and rectal ozone treatment groups (p = 0,011, p = 0,0001). The inflammation values of local ozone treatment group were found to be significantly higher than the control and rectal ozone treatment groups (p = 0,02, p = 0,0001). No statistically significant difference was observed between the other groups (p > 0,05) (Table 2). Mild inflammation was observed in control group and severe inflammation was observed in the local + rectal ozone therapy group (Figure 1. a,b).

Vascular density

The mean vascular density scores of control, local, rectal and local+rectal treatment groups were orderly 1.25±0.46, 2.17±0.75, 2±0 and 3 ± 0 (Table 1). Vascular density values of the local + rectal ozone treatment group were found to be statistically significantly higher than the control, local ozone treatment and rectal ozone treatment groups (p = 0.0001, p = 0.014, p =0,0001). No statistically significant difference was observed between the other groups (p > 0,05) (Table 2). Random vascular structures were and observed in control group severe vascularization was observed in local+rectal ozone treatment group (Figure 2. a,b).

Collagenization

The mean collagenization scores of control, local, rectal and local+rectal treatment groups were orderly 1.50 ± 0.54 , 2.17 ± 0.41 , 2.29 ± 0.49 and 2.57 ± 0.54 (Table 1). The collagenization values of the control group were statistically significantly lower than the local ozone treatment, rectal ozone treatment and local + rectal ozone treatment groups (p = 0,032, p = 0,017, p = 0,006). No statistically significant difference was observed between the other

groups (p> 0,05) (Table 2). Collagen density showed mild staining in lamina propria in the control group. In contrast, local+rectal ozone treatment group had severe collagenization in lamina propria (Figure 3. a,b).

Fibroblast proliferation

The mean fibroblast proliferation scores of control, local, rectal and local+rectal treatment groups were orderly 1.38 ± 0.52 , 2.17 ± 0.41 , 2.29 ± 0.49 and 2.57 ± 0.54 (Table 1). The fibroblast proliferation values of the control group were statistically significantly lower than those of local ozone treatment, rectal ozone treatment and local + rectal ozone treatment groups (p = 0,015, p = 0,008, p = 0,004). No statistically significant difference was observed between the other groups (p> 0,05) (Table 2). Control group had a little fibroblast in lamina propria and many fibroblasts were observed in local+rectal ozone treatment group (Figure 4. a,b).

Epithelization

The mean epithelization scores of control, local, rectal and local+rectal treatment groups were orderly 1.75 ± 0.89 , 2.17 ± 0.41 , 2.29 ± 0.49 and 2.57 ± 0.54 (Table 1). The epithelialization values of the control group were statistically significantly lower than local ozone treatment, rectal ozone treatment and local + rectal ozone treatment groups. (p = 0.042, p = 0.037, p = 0.01). No statistically significant difference was observed between the other groups (p> 0,05) (Table 2). Epithelization was observed as a thin layer in control group and full layer epithelization was observed in local+rectal ozone treatment group (Figure 5. a,b).



Table 1: Mean inflammation, vessel density, collagen density, fibroblast proliferation and epithelization scores of groups

	Inflammation	Vessel Density	0	Fibroblast Proliferation	Epithelization
	Mean±SD	Mean±SD	Mean±SD	Mean±SD	Mean±SD
Control Group	$1,38\pm0,52$	$1,25\pm0,46$	$1,50\pm0,54$	$1,38\pm0,52$	1,75±0,89
Local Ozone Treatment	2 ± 0	2,17±0,75	2,17±0,41	2,17±0,41	2,17±0,41
Rectal Ozone Treatment	1 ± 0	2±0	2,29±0,49	2,29±0,49	2,29±0,49
Local + Rectal Ozone Treatment	2,14±0,38	3±0	2,57±0,54	2,57±0,54	2,57±0,54

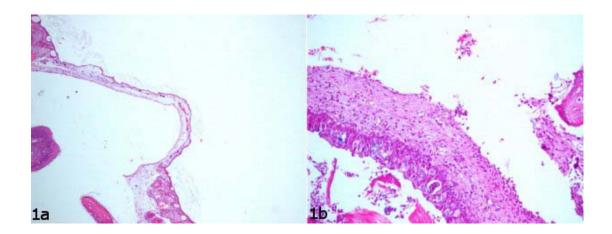


Figure 1: a,b. Mild inflammation (a) was observed in control group and severe inflammation (b) was observed in the local + *rectal ozone therapy group.*

Table 2: Comparison of inflammation, vessel density, collagen density, fibroblast proliferation and epithelization scores of control group and ozone treatment groups

	Inflammation	Vessel Density	Collagen Density	Fibroblast Proliferation	Epitehelization
Dunn's Multiple Comparison Test	P values	P values	P values	P values	P values
Control Group /	0,02	0,023	0,032	0,015	0,042
Local Ozone Treatment					
Control Group /	0,08	0,004	0,017	0,008	0,037
Rectal Ozone Treatment					
Control Group /	0,011	0,0001	0,006	0,004	0,01
Local + Rectal Ozone Treatment					
Local Ozone Treatment /	0,001	0,498	0,626	0,626	0,732
Rectal Ozone Treatment					
Local Ozone Treatment /	0,355	0,014	0,151	0,151	0,234
Local + Rectal Ozone Treatment					
Rectal Ozone Treatment /	0,0001	0,0001	0,298	0,298	0,383
Local + Rectal Ozone Treatment					



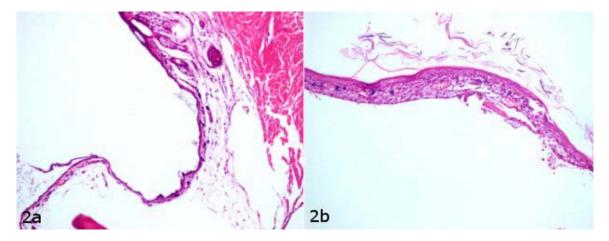


Figure 2: a,b. Random vascular structures (a) were observed in control group and severe vascularization (b) was observed in local+rectal ozone treatment group.

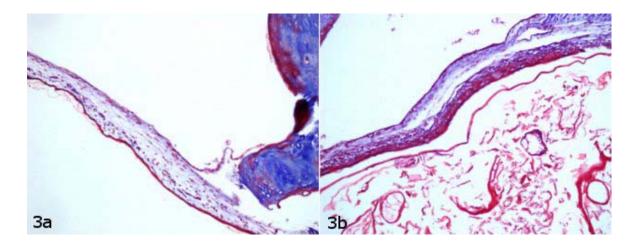


Figure 3: a,b. Control group had mild collagenization (a) in lamina propria and local+rectal ozone treatment group had severe collagenization (b) in lamina propria.

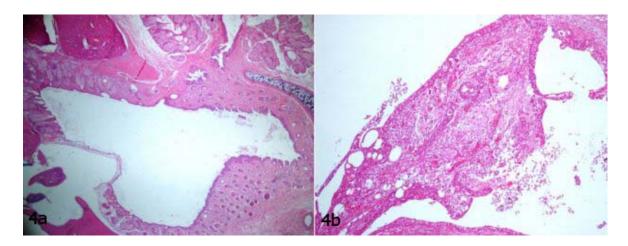


Figure 4: a,b. Control group had a little fibroblast (a) in lamina propria and many fibroblasts (b) were observed in local+rectal ozone treatment group.

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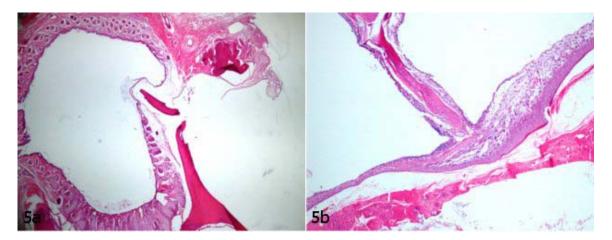


Figure 5: a,b. Epithelization was observed as a thin layer (a) in control group and full layer epithelization (b) was observed in local+rectal ozone treatment group.

DISCUSSION

Wound healing of the tympanic membrane is common in the initial hemostatic and inflammatory stages due to vascular distribution; but it is completely different from other tissues at the proliferation and migration phases. In many wound healing, granulation tissue is formed as a platforma deposit that helps epithelization progress. In typmanic membrane regeneration, rebuilding the of fibrous component follows forming bridges of squamous epithelium layer over wound. Thickened lamina propria contains type 1 and type 3 collagen⁷. The original tympanic membrane contains predominantly type 2 collagen⁸.

When tympanic membrane perforations heal spontaneously with neomembrane, the fibrous layer is deficient. Thus, suboptimal healing may lead to the formation of retraction pockets⁹. In the treatment of ozone, there was a strong middle layer associated with dense collagenization and fibroblast proliferation. Ozone treatment has similar effects with the research of Yeo et al. They also reported PDGF-AA treatment which increases fibroblast number in the middle layer¹⁰. So, ozone therapy provides original tympanic membran formation with 3 layer.

Ozone is a triatomic molecule consisting of 3 oxygen atoms. It has a unstable structure and may convert to molecular oxygen and oxygen atoms. Ozone does not affect by entering into cells. It forms free radicals such as hydrogen peroxide and polyunsaturated fatty acids (PUFAs) in cell membranes. Thus, it increases transcription factors like nuclear factor-kappa B (NF-kB) for synthesis of growth factors and sustaining cell viability¹¹.

Topical ozone application has three main forms; ozone water, ozone oil and ozone gas. Ozone treatment is a cost effective and it has less side effect. As it is used in a wide range of biological applications, it is known that activates the immune system to show antimicrobial effect and increase wound healing¹². On the other hand, ozone treatment is not common in otorhinolaryngology practice. There are clinical trials with ozone therapy which show effectivity of ozone in sudden hearing loss^{13,14}. In contrast, it is not effective in the treatment of tinnitus¹⁵.

Ozone application is beneficial for wound healing. Kim et al. showed the acceleration of healing process in full thickness skin wounds due to administiration of ozone oil¹⁶. In study reported by Erginel et al. the accumulation of collagen in the anastomosis line was observed more in the ozone groups than in the control group after colonic anastomosis¹⁷. In our study, we showed likely that inflammation, fibroblast proliferation, vascular density. collagenization and epithelialization scores were the highest in the local + rectal ozone treatment groups.

Granulocyte Colony Stimulating Factor (G-CSF), Hepatocyte Growth Factor (HGF), Epidermal Growth Factor (EGF), Gelfoam,



Saline irrigation, Ascorbic acid (C vit). Hyaluronic acid were used for experimental studies to accelerate TM repair. Aissa et al. showed gelfoam or G-CSF did not affect to tympanic membrane healing⁹. In the study reported by Güneri et al., there was no statistically significant difference in tympanic membrane thickness, fibroblastic reactions and neovascularization between the 3 groups which are treated with hyaluronic acid, EGF and mitomycin in experimental tympanic membrane perforations¹⁸. In the experimental study reported by Erkilet et al., histopathological evaluation of tympanic membrane healing of PRP group was earlier than the control group, but there was no statistically significant difference between two groups in terms of lamina propria edema, neovascularization fibroblastic reaction and inflammatory cell hyperplasia¹⁹. Similarly, Alan al. found no significant difference et inflammation, epithelial migration and neovascularization on tympanic membrane of between the intraperitoneal ginea pigs pentoxifylline group and intraperitoneal saline group²⁰. In the study reported by Gunes et al, there was no statistically significant difference in terms of inflammation and lamina propria edema between control group and ascorbic acid treatment group²¹. In contrastly, we found statistically increase of vascularization. fibroblast proliferation, and epithelization due to experimental ozone therapy in this study.

Inflammation is the first phase of wound healing and occurs within the first 3-4 days. In our study, severe inflammation reached to the 10th day in ozone treatment groups. Propably, it adversely affect perforation closure due to block passing to another phase. Remodeling phase starts in 3 weeks so our study didn't include remodeling phase. There is a need for studies that include ozone treatment for 3 weeks and evaluates the perforation diameter by microscopic examination.

CONCLUSION

This study showed the contribitions of local and systemic ozone treatment to wound healing by increasing inflammation, fibroblast proliferation, collagenization, vascular density and epithelization in experimental tympanic membrane perforations of rats.

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We have started to this project after the confirmation of Bagcilar Training and Research Hospital Experimental Animal Ethics Board dated 29.03.2016 and numbered 2016/04. There is no conflict of interest and financial disclosure.

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